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Title: Preparation of Sterile PBS for Cell Culture

Version: v5

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SOP History		
Number	Date	Reason for Change
v1	01/01/2013	Original
v2	01/01/2015	Update
V3	01/01/2017	Update
V4	01/01/2019	Update
V5	01/01/2021	Update

1.0 Purpose –

This SOP describes the current procedure for making sterile PBS for cell culture use in Laboratory 248 at the St Andrews School of Medicine (SASoM).

2.0 Scope –

This SOP applies to the staff in the SASoM involved in cell culture work.

3.0 Responsibilities –

All staff involved in cell culture are responsible for ensuring that the methods are followed in accordance with this SOP.

All staff must have read and signed the relevant risk assessment documents before performing this procedure.

4.0 Procedure –

Add 40 PBS tablets (Oxoid; BR0014G) to a 4L beaker.

Make up to 4L with distilled water.

Aliquot into 8 x 500ml Duran bottles and loosely screw on the lids, making sure each bottle has an intact, undamaged and securely-fitting seal around the neck.

Stick a small piece of autoclave tape to the lid of the bottle and write 'PBS' on it.



Autoclave the bottles as per the appropriate SOP.

Remove bottles from autoclave and check that the black bands on the autoclave tape are clearly visible. Allow the bottles to cool before tightening the lids.

Store sterile PBS in the coldroom.

5.0 Personal protection –

A Howie coat must be worn at all times. Gloves as specified in the appropriate COSHH RA.

Thermal gloves must be worn when handling hot items.

6.0 Spillages –

Always clean up any spills immediately after use, only you know what you have spilt and are aware of its hazard.

Spillages should be mopped up with paper towel, disinfected with 70% ethanol and finally washed with detergent.

7.0 Training –

All staff should complete an in house induction to the tissue culture area and be trained in sterile TC techniques before starting any TC work.

8.0 Related documents –

- 8.1 Risk assessments COSHH 04 and RA/BIOL/004



9.0 Approval and sign off –

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