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Title:	Non-Amplified Immunofluorescence using MOUSE and / or RABBIT Primary Antibodies.
Version:	v5
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SOP History		
Number	Date	Reason for Change
V1	30/9/2014	Original
V2	16/02/2015	Minor Amendments
V3	16/02/2017	Additional info re antigen retrieval method and secondary antibody dilution
V4	16/02/2019	Update
V5	16/02/2021	Update

1.0 Purpose –

This SOP describes the current procedure for duplexing Mouse and Rabbit Primary Antibody Immunofluorescence in Laboratory 248 at the St Andrews School of Medicine (SASoM).

2.0 Scope –

This SOP applies to the staff in the SASoM involved with non-amplified Primary Antibody Immunofluorescence. The SOP could be used for single or duplex non-amplified antibody detection / visualisation.

3.0 Responsibilities –

All staff involved in non-amplified antibody immunofluorescence are responsible for ensuring that the methods are followed in accordance with this SOP.

All staff must have read and signed the relevant risk assessment documents before performing this procedure.



4.0 Procedure –

Always put slide(s) in a non-metallic rack.

Antibody detection / visualisation can be carried out using, amongst others, the following secondary antibodies: (i) **Alexa Fluor® 488** (FITC - visualised on either AQUA or Leica DM5500 B microscope), (ii) **Alexa Fluor® 555** (Cy3 - visualised on AQUA or Leica DM5500 B microscope), (iii) **Alexa Fluor 568** (Texas Red - visualised on Leica DM5500 B microscope) or (iv) **Alexa Fluor 647** (Cy5 - visualised on AQUA or Leica DM5500 B microscope).

Visualisation can therefore, in most cases, be performed using both AQUA and the Leica DM5500 B microscope. *Duplexing is possible if required.*

De-wax & Re-hydrate

Always allow any excess fluid to drain from the slide rack before proceeding to the next solution.

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|-------------------|-----------|
| 1. Dewax-Xylene 1 | 5 minutes |
| 2. Dewax-Xylene 2 | 5 minutes |
| 3. Dewax-Xylene 3 | 5 minutes |

During the 3rd xylene dewax stage prepare the antigen retrieval solution; either 1)EDTA-Tris or 2)Citrate based.

- 1) *EDTA-Tris 1 L solution:*
10mM Tris Base = 1.21g
1mM EDTA = 0.37g
Tween = 0.5ml
dH2O = 1L

Add the EDTA and Tris Base to a 1L flask and add the dH2O and mix on magnetic stirrer. Ensure that the pH is 9. Add the Tween.

- 2) *Sodium Citrate/Citric Acid solution for Antigen Retrieval by adding 18mls 0.1M Citric Acid to 82mls 0.1M Sodium Citrate (100mls). Make up to 1 litre with Elga Water (pH6).*

Pour the antigen retrieval solution into the microwave pressure cooker. Screw on the lid and put into the microwave on high power to heat up for 10mins.

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| 4. Rehydration-100% Alcohol | 2 minutes |
| 5. Rehydration-100% Alcohol | 2 minutes |
| 6. Rehydration-80% Alcohol | 2 minutes |
| 7. Rehydration-50% Alcohol | 2 minutes |
| 8. Wash in running tap water (increasing to hot) | 2 minutes |

Antigen Retrieval using microwave pressure cooker

1. Perform antigen retrieval on slides by putting drained slide holder into pre-heated buffer in pressure cooker. Screw on the lid and place the red weight over the vent. Put back into the microwave and heat on high for 8 mins – there should be an audible hissing sound towards the end of the procedure..
2. Using heat resistant gloves and eye protection carefully lift the pressure cooker from within the microwave and place in sink. Place eye protection on. With the heat resistant gloves release the pressure by removing the weight from the top of the cooker. Stand well back immediately once the weight is removed. Do not stand over the pressure cooker during this step.
3. Remove the lid from the pressure cooker using the heat resistant gloves and $\frac{3}{4}$ fill with cold water. Allow the slides to cool down for 20mins.

Antigen Retrieval using Instant pot electric pressure cooker –Please read and sign SASoM-EQUIP/100.v1

1. Make sure the pressure release handle and float valve are unobstructed and clean, and that the sealing ring is properly inserted.
2. Fill the inner pot with 1 litre of antigen retrieval solution (either Sodium Citrate buffer or Tris EDTA buffer).
3. Close the lid. Make sure that the pressure release handle is pointing to the “Sealing” mark on the lid (Figure 1).

**Figure 1 Pressure release handle**

4. Press “Manual” button, and press the “+” or “-” to change the time to 1 min. In 10 seconds, the Instant pot will go into preheating cycle (display showing “On”)(Figure 2).

**Figure 2 Control panel**



5. Once working pressure is reached, which make take a few minutes or up to 10-13mins, the countdown timer will begin. When the countdown is finished, the Instant pot will beep and automatically switch into the “Keep Warm” mode.
6. Turn the pressure release handle to the “Venting” position to let out steam until the float valve drops down (Figure 1). Open the lid with care. **When releasing steam, always wear a pair of thermal gloves and a safety goggles.**
7. Add the rehydrated slides into the inner pot and close the lid. Make sure that the pressure release handle is pointing to the “Sealing” mark on the lid (Figure 3).
8. Press “Manual” button, and press the “+” or “-“ to change the time to 5 min. In 10 seconds, the Instant pot will go into preheating cycle (display showing “On”).
9. Once working pressure is reached, which make take a few minutes or up to 10-13mins, the countdown timer will begin. When the countdown is finished, the Instant pot will beep and automatically switch into the “Keep Warm” mode.
10. Press “Cancel” and Turn the pressure release handle to the “Venting” position to let out steam until the float valve drops down (Figure 1). Open the lid with care.
11. Take the whole inner pot out and add the running tap water and cool the slides down.

Blocking

1. Transfer slide(s) into coplin jar and then wash in PBS/Tween 20 (500mls PBS +500µl Tween 20) for 5 minutes.
2. Treat sections in 3% Hydrogen peroxide (10ml of 30% H₂O₂ stock solution plus 90mL Elga H₂O) for 5 minutes. Store hydrogen peroxide in the fridge.
3. Wash slides in PBS/T for 5 minutes. Blot off excess solution from edge of slides onto tissue. Draw round individual sections on slide with hydrophobic Immedge pen.
4. Block with Dako Total Protein blocking solution (taken straight from fridge or keep on ice) for 10 minutes by dropping solution onto individual sections (as indicated on solution data sheet).
5. Blot off Blocking solution on tissue.

Primary Antibody Incubation

1. Add the two primary antibodies diluted at appropriate concentrations in Dako antibody diluent (previously optimised via IHC). It is important that the two primary antibodies are from a different host species.
2. Incubate either 30min at room temperature or at 4°C overnight and then rinse in 0.1% PBST 2 x 5 minutes.

Antibody visualisation

1. Prepare a 1 in 100 dilution of the appropriate Alexa Fluor® secondary antibody(s) in Dako antibody diluent. *There are both mouse and rabbit variants of each Alexa Fluor so every combination can be used for duplexing.*



2. Incubate slides in the dark for 30min at room temperature. If there are problems with visualisation, the slides can alternatively be incubated in the HybEZ oven (37°C) for 45 minutes.
3. Rinse in 0.1% PBST 2 x5 minutes

Counterstaining and Coverslipping

1. Blot off excess PBS onto tissue. Apply 40 µl (22x40mm coverslip) Prolong Gold anti-fade reagent with DAPI (Invitrogen, P36931), nuclear visualisation media, onto the sample and gently lower the coverslip over the tissue, ensuring no air bubbles are present. (For the smaller coverslips (22x26mm), apply 20µl).
2. Refer to TSA MSDS sheet for guidance on removal of coverslip to remount, if necessary.
3. Tack down coverslip at 4 corners with small quantity of nail varnish to prevent accidental movement of coverslip.
4. Let the mounted slide dry for at least 24hrs in horizontal position, in the dark, to 'cure'.
5. After slides are 'cured', seal the coverslips with nail polish. Allow to dry in horizontal position and place in slide box for long term storage

5.0 Personal protection –

A Howie coat must be worn at all times. Gloves as specified in the appropriate COSHH RA.

6.0 Spillages –

Always clean up any spills immediately after use, only you know what you have spilt and are aware of its hazard.

Spillages should be mopped up with paper towel, disinfected with 70% ethanol and finally washed with detergent.

7.0 Training –

All staff should under go training in this technique before performing the procedure.



8.0 Related documents –

- 8.1 Risk assessments COSHH 19
- 8.2 SOP SASoM/METHOD/024
Dewaxing and Rehydration of Paraffin Embedded Sections
- 8.3 SOP SASoM/EQUIP/015
Use of the Pressure Cooker for Antigen Retrieval
- 8.4 SOP SASoM-EQUIP/100.v1
Use of Instant pot electric pressure cooker for antigen retrieval

9.0 Approval and sign off –

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